

Supporting information for:

Proton-coupled hopping in Ru-modified
P. aeruginosa azurin

Jeffrey J. Warren, Oliver S. Shafaat, Jay R. Winkler and Harry B. Gray*

Beckman Institute
California Institute of Technology
Pasadena CA 91125
USA

hbgray@caltech.edu

Contents:

1.	Mass spec characterization of Ru-labeled proteins	S2
2.	pK _a of YOH107 and YOH124.....	S4
3.	Kinetics traces at different buffer concentrations	S5
4.	Kinetics traces in different buffers	S7

1. Mass spec characterization of Ru-labeled proteins

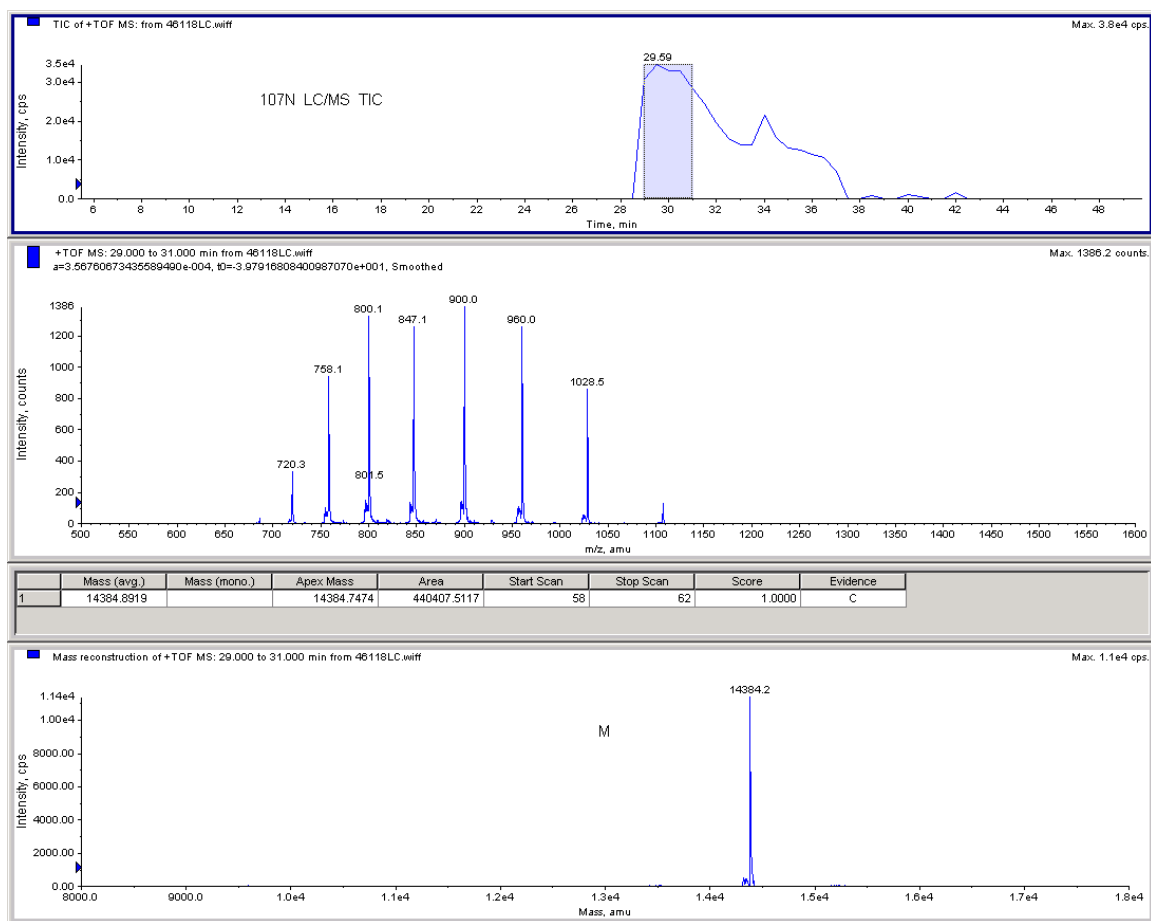


Figure S1. Liquid chromatography-mass spec for RuH107YOH109 azurin. Expected mass: 14388 Da; observed mass: 14384 Da.

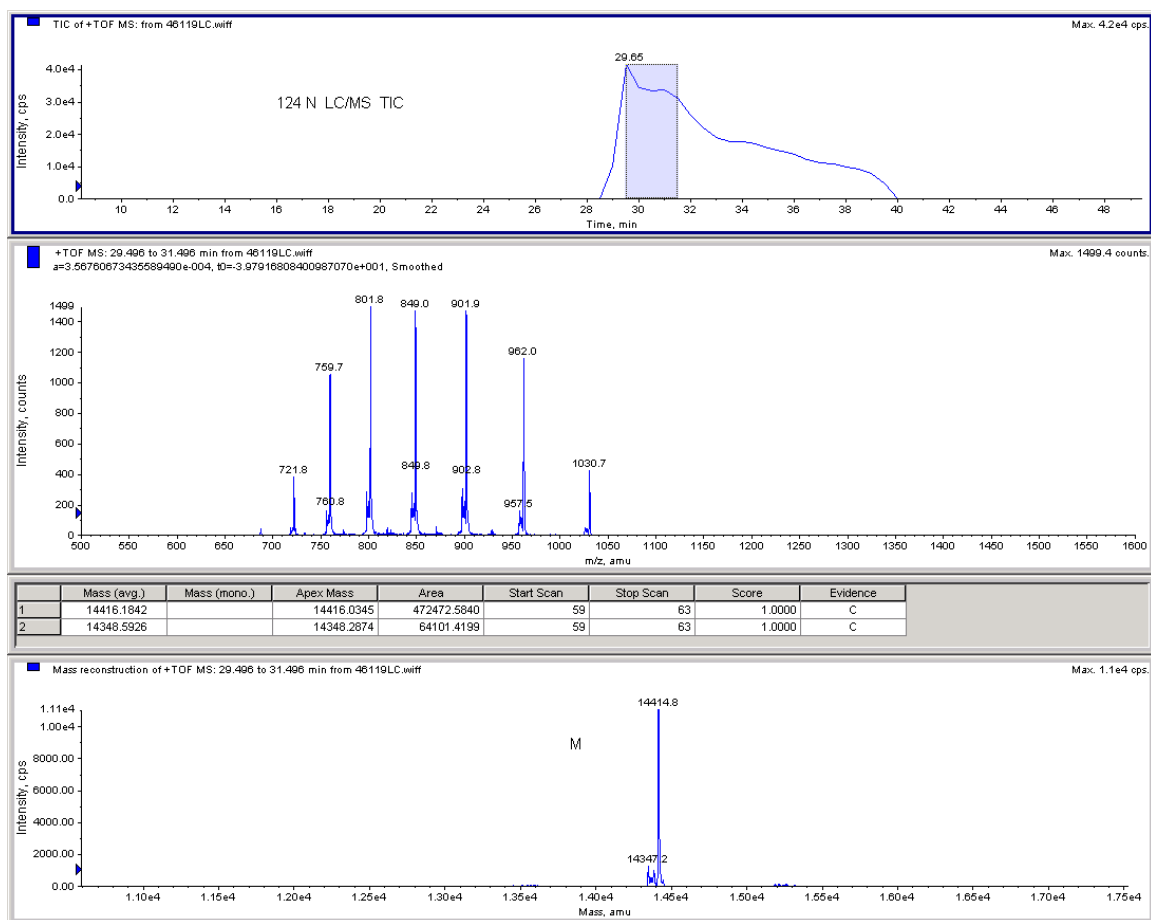


Figure S2. Liquid chromatography-mass spec for RuH124YOH122 azurin. Expected mass: 14418 Da; observed mass: 14415 Da. The MS for RuH126YOH122 is identical, as the proteins have the same expected masses.

2. pK_a of YOH107 and YOH124

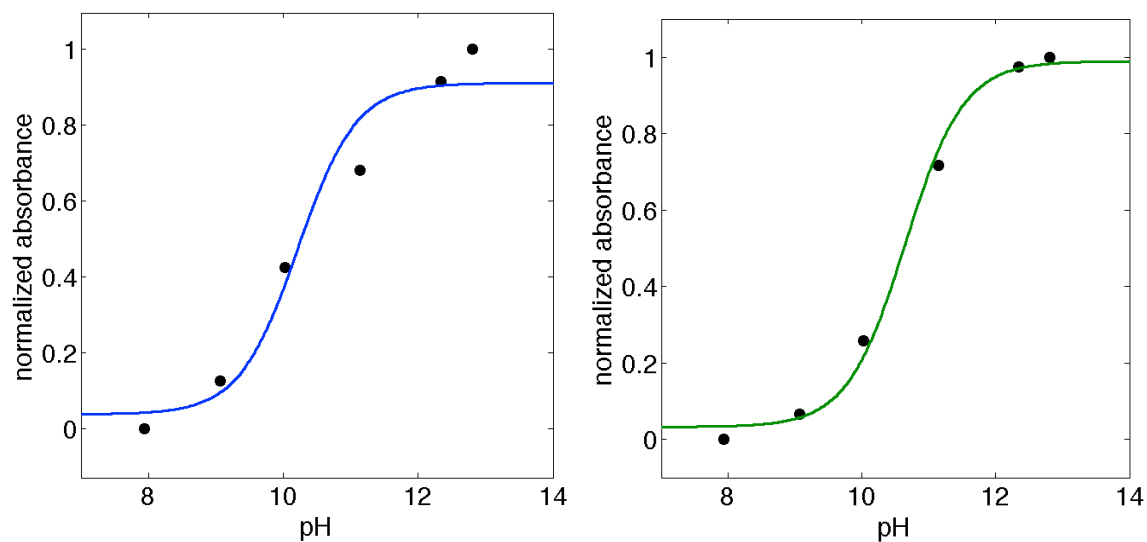


Figure S3. Titration data for H107YOH109Cu^{II} (left) and H122YOH124Cu^{II} (right) azurins. Data were collected in 250 mM sodium borate + 100 mM NaCl at 293 K. The fits (see main text) give $pK_a(\text{YOH109}) = 10.2 \pm 0.2$ and $pK_a(\text{YOH122}) = 10.6 \pm 0.1$.

3. Buffer concentration dependence kinetics traces.

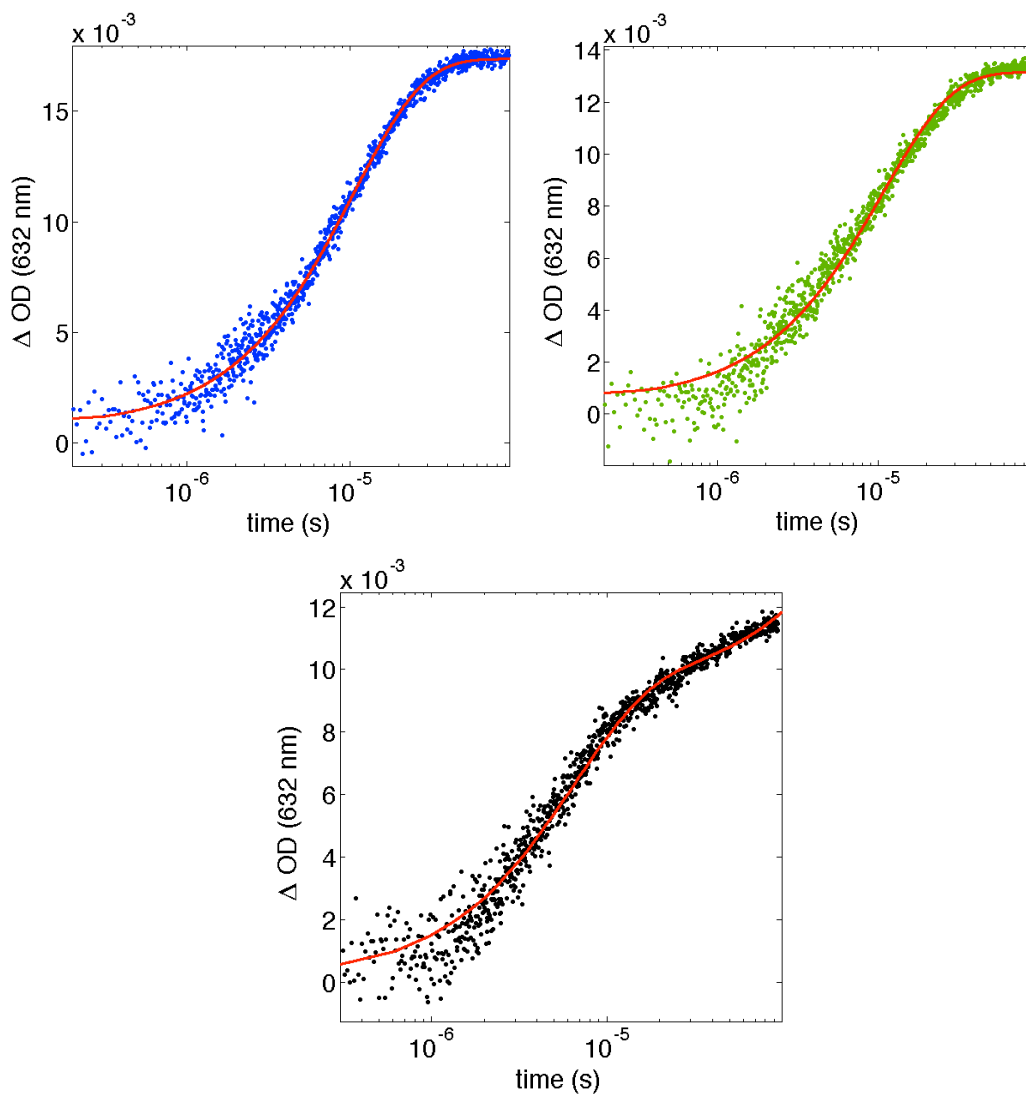


Figure S4. Kinetics traces for RuH124YOH122 Cu^I oxidation in imidazole buffer at pH 8. (A) 250 mM (as in the main text). (B) 100 mM. (C) 10 mM with a biexponential fit. $k_1 = 1.6 \times 10^5$ and $k_2 = 2.5 \times 10^3$.

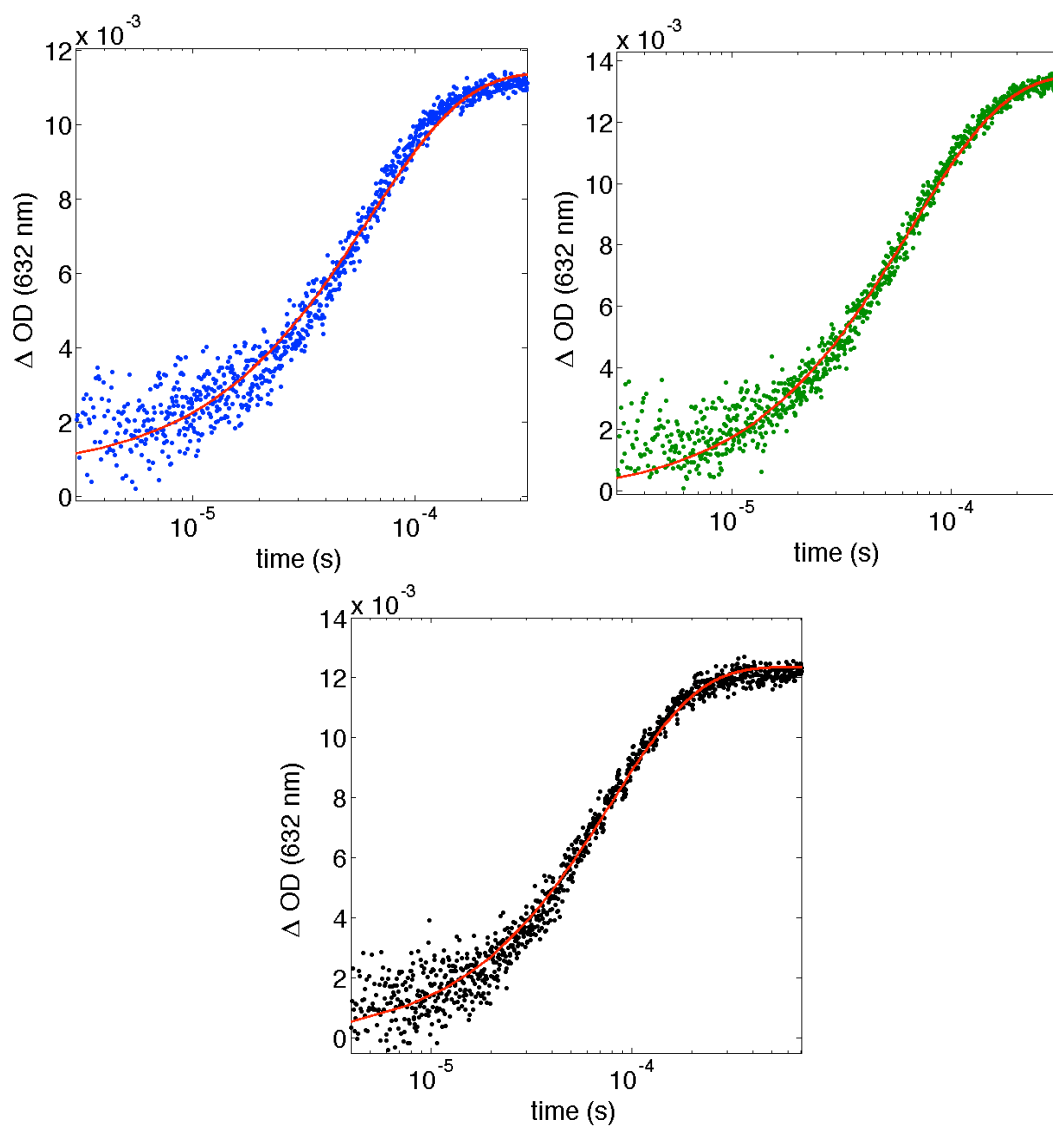


Figure S5. Kinetics traces for RuH107YOH109 Cu^I oxidation in imidazole buffer at pH 8. (A) 250 mM (as in the main text). (B) 100 mM. (C) 10 mM.

4. Kinetics traces in different buffers

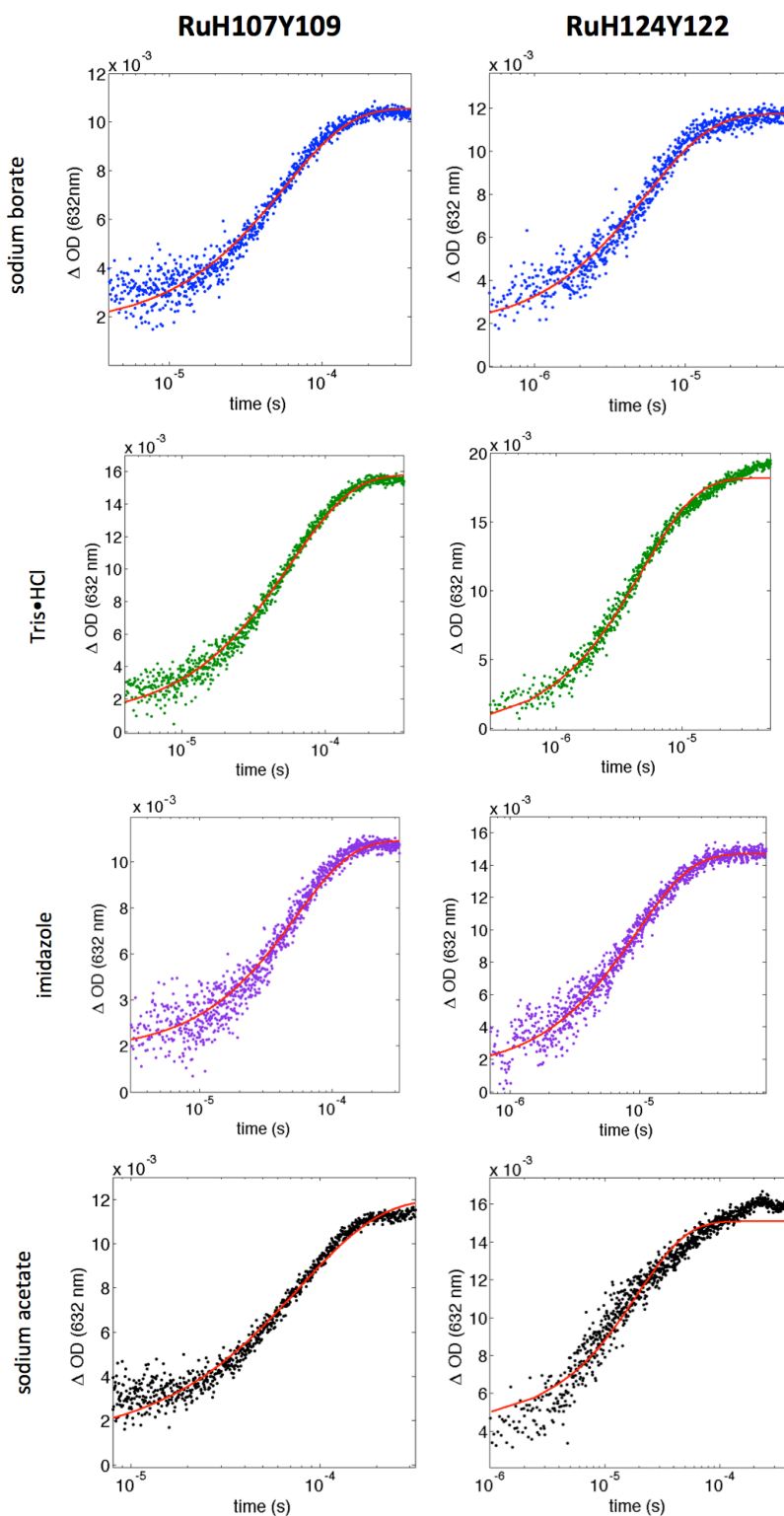


Figure S6. Kinetics traces for Cu^{I} oxidation in different buffers (all 250 mM, pH 8). The left column shows data for RuH107Y109 azurin and the right column shows the data for RuH124Y122 azurin. Buffer pK_{a} s are: 9.14 (borate); 8.07 (Tris); 7.05 (imidazole); 4.76 (acetate).